

# Amylase and Protease Activities of Microorganisms Isolated from Cassava Wastewater Contaminated Soil

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# ABSTRACT

Agro waste management is one of the problems faced by humans in their environment especially in many developing nations. In Nigeria, several agro wastes are generated in enormous quantities including cassava processing wastewater. Nigeria is the world-leading producer of cassava and it has been estimated that 5.142 million tonnes of cassava wastewater are discharged into the Nigerian environment of which 45–65% could be recovered. It has also been projected that the wastewater could increase significantly before 2035. The cassava wastewater is often discharged without any formal treatment methods by gari processors in many areas of cassava producing communities in Nigeria. Untreated cassava wastewater leads to environmental degradation and loss of biodiversity due to its lethal characteristics. Cassava wastewater can be harnessed through its use in biotechnological advances especially enzyme production. This study was designed to assess the amylase and protease activities of some microbial isolates (Escherichia coli, Staphylococcus aureus, Pseudomonas, Bacillus, Micrococcus, Proteus, and Enterobacter species viz: bacteria, and Saccharomyces cerevisiae, Aspergillus, Mucor, Penicillium and Rhizopus species viz: fungi) from cassava wastewater contaminated soil. The protease and amylase activities from the isolates were carried out using standard microbiological processes. The results showed that all the microbes have amylase and protease activities except for Escherichia coli and Enterobacter species, and Proteus and Micrococcus species showed only amylase and protease activity, respectively. These enzymes (amylase and protease) can be applied in numerous industrial sectors in the nation's economy. As such, microbes from cassava wastewater contaminated soil could be obtained and used for the production of useful enzymes with biotechnological potentials.

**Keywords:** Amylase, Cassava mill effluents, Enzymes, Microorganisms, Protease

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### Introduction

Enzymes are biomolecules produced by living organisms that help to accelerate various biological/ biochemical reactions that are pivotal in sustaining human life. Enzymes are efficient and specific biocatalysts that can speed up reaction rates several times faster than normal chemical reactions both in and out of the cell. Enzymes are present in all living things ranging from plants to animals including microorganisms. However, in plants and animals enzymes are produced in smaller quantities and are rarely exploited for industrial uses whereas microbial enzymes have found widespread uses in industries. This is because microbial enzymes are more active and stable, easier to handle, rapid multiplication and produce high yield, cheap to produce and are more susceptible to genetic manipulation.

Industrial uses of microbial enzymes are gaining more attention due to their eco-friendly and non-toxic nature. Other special characteristics such as thermotolerance, stability over different ranges of temperature and pH, and other harsh conditions have made microbial enzymes very useful in different commercial industries. Enzymes are generally biosynthesized from microbes that have advanced in biotechnology such as recombinant DNA technology, protein engineering, and metagenomics which have revolutionized the commercialization of enzymes with new activities and adaptabilities further increasing their use in different industrial processes.<sup>1</sup> Enzymes are useful in the manufacturing sector such as textile, food, pharmaceuticals, cleaning agents, biofuel, etc. These applications make microbial enzyme market very competitive. One of the most useful enzymes in industries is hydrolases which catalase the breakdown of molecules. Enzymes such as amylase and protease have been widely studied with the advent of biotechnology due to their extracellular hydrolytic properties.

Amylase is used as an additive in several production processes including pharmaceutical, food, fermentation, textile, paper,<sup>2-4</sup> detergents,<sup>3</sup> brewing and distilling industries, fine-chemical, clinical, medicinal and analytical chemistries.<sup>2</sup> Similarly, amylase has also found applications in breweries and biofuel industries.<sup>5</sup> Alpha-Amylase is also a useful conversion of starches into oligosaccharides.<sup>3</sup> Starch-converting enzymes are useful in the production of maltodextrin, glucose, and fructose syrups.<sup>3</sup> Proteases, on the other hand, have found applications in detergent production, food enzymes (used for baking, brewing, cheese production) meat tenderization, leather, textiles and fabrics<sup>6,7</sup> pulp, paper making industry, bioremediation processes<sup>7</sup> production of protein hydrolysates, solubilization of keratin materials to convert waste materials into useful products, in silver recovery from conventional gelatincontaining photographic film, liquefaction of organic waste and it can even be consumed by humans and animals and used as therapy during thrombosis and cancer treatment.<sup>6</sup>

Cassava wastewater contains groups of microorganisms including acid formers and hydrocarbon utilizers.<sup>8</sup> Like palm oil mill effluents, cassava wastewater is capable of producing lipolytic microbes including bacteria and fungi (mold and yeasts). The ability of these microbes to thrive in cassava wastewater could be associated with the availability of basic minerals in the wastewater. The microorganisms found in cassava wastewater could be converted into useful products of high demand. In the traditional setting in Nigeria, cassava wastewater is converted into starch. Cassava wastewater is rich in cellulose and hemicellulose. So microbes can mineralize the nutrients found in starches substrates such as cassava waste water with the action of additives to produced enzymes such as amylase, protease, cellulase, lipase, etc. Some of these enzymes such as cellulase,<sup>9</sup> amylase,<sup>2,9,10</sup> and protease<sup>11-16</sup> can be produced from some microorganisms. Adejuwon et al.17 reported that Candida albicans produces an optimum protease at a temperature and pH of 30°C and 7.0 respectively. Therefore, this paper focussed in assessing protease and amylase potentials of microbes isolated from cassava wastewater contaminated soil.

#### **Materials and Methods**

#### Data Source

The microbial isolates (*Escherichia coli, Staphylococcus aureus, Pseudomonas, Bacillus, Micrococcus, Proteus,* and *Enterobacter* species for bacteria, and *Saccharomyces cerevisiae, Aspergillus, Mucor, Penicillium* and *Rhizopus* species for fungi) used for this study were previously isolated from cassava wastewater contaminated soil by Izah and Aigberua.<sup>18</sup>

#### **Amylase Assessment**

Potatoes extract was incorporated into Nutrient Agar (for the determination of amylase producing bacteria) and Sabouraud Dextrose Agar (for the determination of amylase producing fungi) in the ratio of 40:60. The pure isolates were aseptically inoculated unto the respective agar plates and incubated for 24-48 hours (for bacteria) and 3-4 days (for Fungi/yeast) at 37°C. Then after, the agar plates were flooded with iodine solution, and a clear zone on the plates exposed to Gram's iodine is an indication of amylase producing ability of the isolates.<sup>9,19-21</sup>

#### **Protease Assessment**

Distilled water and liquid peak milk bought from a commercial shop in port Harcourt, Nigeria was incorporated into double strength media (Nutrient Agar used for protease producing bacteria, and Sabouraud Dextrose Agar for protease producing fungi) in the ratio of 35:65. Also, K<sub>2</sub>HPO<sub>4</sub>

2.0, peptone 5.0, gelatin 15.0 was added to the agar.<sup>22,23</sup> *fermenti* as  $\alpha$ -amylase price were aseptically inoculated unto the respective cassava peels at an incuration of organisms and subsequently similarly. Senthilkumar

The pure were aseptically inoculated unto the respective agar plates for both groups of organisms and subsequently incubated at 37°C for 24-48 hours for bacteria and 3-4 days for fungi. The agar plates were flooded with mercury chloride.<sup>22,23</sup> A clear zone at the end of the incubation period around each isolate/colonies indicated protease production.<sup>12,22,23</sup> which was quantitatively expressed.

#### **Results and Discussion**

Table 1, shows the amylase and protease activities of microorganisms isolated from cassava wastewater contaminated soil. *Staphylococcus aureus, Pseudomonas aeruginosa, Bacillus* species (bacteria), and *Saccharomyces cerevisiae, Aspergillus, Mucor, Penicillium* and *Rhizopus* species (fungi) are both amylase and protease producers. While *Escherichia coli* and *Enterobacter* species did not produce amylase or protease. *Proteus* species showed a slight amylase ability and no protease potential. Also, *Micrococcus* showed protease activity and no amylase potential.

Table I.Amylas	e and protease	activity o	of microbes
found in cassa	va wastewater	contamir	nated soil

Organisms	Amylase activity	Protease activity
Bacteria		
Pseudomonas aeruginosa	+	+
Escherichia coli	-	-
Staphylococcus aureus	+	+
Micrococcus species	-	+
Proteus species	+	-
Enterobacter species	-	-
Bacillus species	+	+
Fungi (mould and yeast)		
Aspergillus species	+	+
Saccharomyces cerevisiae	+	+
Penicillium species	+	+
Mucor species	+	+
Rhizopus species	+	+

(+ = ability; and - = no ability)

The microbes found to possess amylase producing potentials in this study has some similarity with the findings of previous reports. For instance, Oboh<sup>24</sup> studied cassava mill effluents and reported that pure strains of *Saccharomyces cerevisiae* along with *Lactobacillus delbruckii* and *Lactobacillus coryniformis* for 72 hours are capable of producing amylase. Akpomie et al.<sup>20</sup> reported *Bacillus subtilis, Bacillus megaterium, Corynebacterium kutseri* and *Lactobacillus*  *fermenti* as  $\alpha$ -amylase producing microorganisms from cassava peels at an incubation temperature of 26-37°C. Similarly, Senthilkumar et al.25 reported that Bacillus species have amylase activity using cassava as a substrate. The result presented here is also similar to other noncassava substrates studies. Ohimain et al.5 reported that Staphylococcus aureus, Bacillus, Pseudomonas, Penicillium, Fusarium, Mucor, Candida species and Aspergillus niger as amylase producing microorganisms from palm oil mill effluents. Kongkiattikajorn<sup>26</sup> reported that amylase and glucoamylase can be produced from Saccharomyces diastaticus. Ruban et al.<sup>27</sup> reported that Bacillus subtilis and Aspergillus niger isolated from soga effluents are capable of producing amylase. Sanni et al.<sup>28</sup> reported Lactobacillus plantarum and Lactobacillus fermentum as amylase producing microbes from different traditionally fermented foods in Nigeria. Ikram-Ul-Haq et al.<sup>29</sup> screened fungi isolates and reported that Saccharomyces cerevisiae and Asperaillus niger are  $\alpha$ -amylase producers. Abu et al.<sup>30</sup> reported that a mixture of Aspergillus niger and Saccharomyces cerevisiae grew on sorghum pomace can hydrolyze its starch content to form amylase. Sugita et al.<sup>31</sup> studied amylase production from the intestinal microflora of freshwater fish and reported that Pseudomonas, Chrostridium, Aeromonas, Bacteriodaceae species can produce amylase. Oseni and Ekperigin<sup>32</sup> reported that Streptococcus feacalis, Escherichia fruendi, Bacillus megatarium, Kurthia species, Erwinia amylovora, Lactobacillus acidophilus, Proteus vulgaris, and Proteus mirabilis have  $\alpha$ -amylase activity in an incubation temperature range of 30-60°C. Alariya et al. <sup>19</sup> reported that Pseudomonas fluorescens, Bacillus subtilus, Escherichia coli, and Serratia marscens possesses amylase activity. Escherichia coli are not known to be amylase producers, but when an amylase producing gene from other microbes are inserted into the Escherichia coli, it could be transformed into an abundant amylase producer.33

The protease produced from microorganisms found in cassava wastewater contaminated soil is comparable to the report from other substrates. Authors have reported that different microbes possess protease activity depending on the minerals/ nutrients available. For instance, Mucor circinelloides produces protease using glucose as a substrate,<sup>34</sup> Fusarium culmorum, F. avenaceum and F. oxysporum using a mineral-protein medium and a cell wall medium.<sup>35</sup> Some other microbes that have been reported to possess protease activity include Aspergillus,14 Lactobacillus, <sup>15</sup> Bacillus, Staphylococcus<sup>16</sup> and Mucor species.<sup>13</sup> Odu and Akujobi<sup>36</sup> reported that *Micrococcus luteus* and Bacillus species isolated from an abattoir environment produces protease which reaching optimum at 37°C and 7.0 for temperature and pH, respectively. The authors further reported that Bacillus, Pseudomonas, Halomonas, Arthrobacter and Serratia as important protease producing

bacteria. Vermelho et al.<sup>37</sup> reported that *Pseudomonas* aeruginosa, Micrococcus luteus and Serratia marcescens have proteolytic activities. Chaturvedi et al.<sup>38</sup> reported that Fusarium, Curvularia, Aspergillus and Mucor species as good sources of extracellular protease production. Arsenijevic et al.<sup>39</sup> reported that *Candida* species have protease activity. Sharmin et al.<sup>40</sup> reported that *Lactobacillus* species isolated from the rumen have proteolytic properties producing at an optimum temperature, pH and incubation period of 37°C, 8.0 and 2 days, respectively. Raja et al.<sup>41</sup> reported that Penicillium species have proteolytic activity on Skimmed Milk Agar Medium. Mishra<sup>42</sup> reported that Lactobacillus delbrueckii has been known for their protease producing activity and they further stated that it can mutate to give biochemical properties similar to Neisseria flavescens. Owoseni and Onilude<sup>43</sup> studied the protease production from Enterobacter species and Escherichia coli isolated from processed foods and reported that acid-stable protease, metalloprotease, and not serine-proteases reaches the maximum at 50°C for both organisms. Microorganisms and its products particularly enzymes have been applied in different sectors. The extracellular enzyme activity of cassava wastewater could be attributed to the mineral and nutrient available in the effluents. Also, most of the microbes under study possess amylase and protease activity which is dependent on the incubation period and temperature, pH of the medium, source and biochemical characteristics of the isolates and choice of media used for investigation.

## Conclusion

This study assessed the microbial isolates from cassava wastewater contaminated soil for the production of amylase and protease. The majority of the isolates including Staphylococcus aureus, Pseudomonas aeruginosa, Bacillus species (bacteria), and Saccharomyces cerevisiae, Aspergillus, Mucor, Penicillium and Rhizopus species (fungi) are amylase and protease producers. Escherichia coli and Enterobacter species cannot produce amylase and protease. While Proteus species showed slight amylase ability and no protease potential. Similarly, Micrococcus species only produced protease. As such, cassava wastewater contaminated soil could be a possible source of microbes for the production of extracellular enzymes needed by several industrial and biotechnology industries. By so doing, the attendant environmental impacts associated with the wastewater can be reduced.

#### References

 Wong CF, Rahman RNZRA, Salleh AB et al. Characterization of Recombinant Organic Solvent Tolerant Proteases. In: Molecular and Structural Biology of New Lipases. Rahman RNZRA, Salleh AB, Basri M (editors). 2013; 69 -88.

- Pandey A, Nigam P, Soccol CR et al. Advances in microbial amylases. *Biotechnol Appl Biochem* 2010; 31: 135-152
- de Souza PM, Magalhães P, de O. Application of microbial α-amylase in industry - a review. Brazilian Journal of Microbiology 2010; 41: 850-861
- 4. John J. Amylases-bioprocess and potential applications: a review. *International Journal of Bioinformatics and Biological Sciences* 2017; 5: 41-50.
- Ohimain El, Izah SC, Jenakumo N. Physicochemical and Microbial screening of palm oil mill effluents for amylase production. *Greener Journal of Biological Sciences* 2013; 3(8): 314-325.
- Ward OP. Proteases. Comprehensive Biotechnology, 3<sup>rd</sup> edition. 2011; 3: 604-615. https://doi.org/10.1016/ B978-0-444-64046-8.00187-7
- Jabalia N, Mishra PC, Chaudhary N (2014). Applications, Challenges and Future Prospects of Proteases: An Overview. Journal of Agroecology and Natural Resource Management 1(3): 179-183
- Ohimain EI, Daokoru-Olukole C, Izah SC et al. Microbiology of palm oil mill effluents. *Journal of Microbiology and Biotechnology Research* 2012; 2(6): 852-857.
- 9. Arotupin DJ. Evaluation of microorganisms from cassava waste water for production of amylase and cellulose. *Research Journal of Microbiology* 2007; 2(5): 475-480.
- 10. Oshoma CE, Imarhiagbe EE, Ikenebomeh MJ et al. Nitrogen supplements effect on amylase production by Aspergillus niger using cassava whey medium. *African Journal of Biotechnology* 2010; 9(5): 682-686.
- 11. Santhi R. Microbial production of protease by Bacillus cerus using cassava waste water. *European Journal of Experimental Biology* 2014; 4(2): 19-24.
- Okorie PC, Olasupo NA. Growth and extracellular enzyme production by microorganisms isolated from Ugba-an indigenous Nigerian fermented condiment. *African Journal of Biotechnology* 2013; 12(26): 4158-4167.
- 13. Alves MH, de Campos-Takaki GM, Porto ALF et al. Screening of *Mucor* spp. for the production of amylase, lipase, polygalacturonase and protease. *Brazilian Journal of Microbiology* 2002; 33: 325-330.
- 14. Shivakumar S. Production and characterization of an acid Protease from a local *Aspergillus* Sp. by Solid substrate fermentation. *Archives of Applied Science Research* 2012; 4(1): 188-199.
- 15. Sasaki M, Bosman BW, Tan PST. Comparison of proteolytic activities in various lactobacilli. *Journal of Dairy Research* 1995; 62(4): 601-610.
- Ramalakshmi N, Narendra D, Ramalakshmi M et al. Isolation and characterization of protease producing bacterial from soil and estimation of protease by

spectrophotometer. *The Experiment, International Journal of Science and Technology* 2012; 1(1): 1-7

- 17. Adejuwon AO, Lamidi MT, Akintobi OA et al. Production of Protease from a Strain of *Candida albicans*: Flour as Growth Substrate. *Report and Opinion* 2013; 5(6): 23-26.
- Izah SC, Aigberua AO. Assessment of Microbial Quality of Cassava Mill Effluents Contaminated Soil in a Rural Community in the Niger Delta, Nigeria. EC Microbiology 2017; 13(4): 132-140.
- 19. Alariya SS, Sethi S, Gupta S et al. Amylase activity of a starch degrading bacteria isolated from soil. *Archives of Applied Science Research* 2013; 5(1): 15-24.
- 20. Akpomie OO, Akponah E, Okorawhe P. Amylase production potentials of bacterial isolates obtained from cassava root peels. *Agricultural Science Research Journals* 2012; 2(2): 95-99.
- 21. Benson HJ. Microbiological Applications: Laboratory Manual in General Microbiology/complete version, 5th edition. McGaraw-Hill, New York. 2002.
- 22. Alnahdi HS. Isolation and screening of extracellular proteases produced by new Isolated Bacillus sp. *Journal of Applied Pharmaceutical Science* 2012; 2(9): 071-074
- 23. Sony IS, Potty VP. Isolation and Identification of Protease Producing Bacteria from Food Processing Industries. International Journal of Current Microbiology and Applied Science 2016; 5(3): 181-189
- 24. Oboh G. Isolation and characterization of amylase from fermented cassava (*Manihot esculenta* Crantz) wastewater. *African Journal of Biotechnology* 2005; 4(10): 1117-1123.
- Senthilkumar PK, Uma C, Saranraj P. Amylase Production by *Bacillus* sp. Using Cassava as Substrate. *International Journal of Pharmaceutical & Biological Archives* 2012; 3(2): 300-306
- 26. Kongkiattikajorn J. Production of Amylase from *Saccharomyces diastaticus* sp, Hydrolysis of Cassava Pulps for Alcohol Production. *Journal of Agricultural Science and Technology* 2012; 2: 909-918.
- 27. Ruban P, Sangeetha T, Indira S. Starch Waste as a Substrate for Amylase Production by Sago Effluent Isolates *Bacillus subtilis* and *Aspergillus niger. American-Eurasian J Agric & Environ Sci* 2013; 13(1): 27-31.
- 28. Sanni AI, Morlon-Guyot J, Guyot JP. New efficient amylase-producing strains of Lactobacillus plantarum and *Lactobacillus* fermentum isolated from different Nigerian traditional fermented foods. *Int J Food Microbiol* 2002; 72(1-2): 53-62.
- 29. Ikram-UI-Haq, Abdullah R, Ashrat H et al. Isolation and screening of fungi for biosynthesis of alpha amylase. *Biotechnology* 2002; 1(2-4): 61-66.
- 30. Abu AB, Ado SA, James DB. Raw starch degrading amylase production by mixed culture of *Aspergillus*

*niger* and *Saccharomyces cerevisae* grown on sorghum pomace. *African Journal of Biotechnology* 2005; 4(8): 785-790.

- 31. Sugita H, Kawasaki J, Deguchi Y. Production of amylase by the intestinal microflora in cultured freshwater fish. *Letter in Applied Microbiology* 1997; 24: 105-108.
- 32. Oseni OA, Ekperigin MM. Isolation and activity of alpha amylase from selected bacteria strains in the forest soil. *Global Journal of Bioscience and Biotechnology* 2013; 2(1): 17-20.
- 33. Yin Q, Xue G, King A et al. Transformed Escherichia coli with amylase gene from Bacillus subtilis. Recent advances in animal nutrition, RAAN conference proceedings. 2001.
- Andrade VS, Sarubbo LA, Fukushima K et al. Production of extracellular proteases by mucor circinelloides using d-glucose as carbon source/ substrate. *Brazilian Journal* of Microbiology 2002; 33: 106-110
- 35. Urbanek H, Yirdaw G. Acid proteases produced by Fusarium species in cultures and in infected seedlings. *Physiological Plant Pathology* 1978; 13(1): 81-87
- 36. Odu NN, Akujobi CO. Protease Production Capabilities of *Micrococcus luteus* and *Bacillus* species Isolated from Abattoir Environment. *Journal of Microbiology Research* 2012; 2(5): 127-132.
- Vermelho AB, Meirelles MML, Lopes A et al. Detection of Extracellular Proteases from Microorganisms on Agar Plates. *Mem Inst Oswaldo Cruz,* Rio de Janeiro, 1996; 91(6): 755-760
- Chaturvedi S, Pathak S, Upadhyay R et al. Comparative Study of Dermatophytic Fungi for Extra Cellular Proteases Efficacy. *Research and Reviews: Journal of Microbiology and Biotechnology* 2013; 2(3): 66-77.
- 39. Arsenijevic VA, Arsovic N, Dzamic A et al. Protease activities of *Candida* spp. isolated from immunocompetent patients with Otomycosis. *Jugoslov Med Biohem* 2004; 23: 171-174.
- 40. Sharmin S, Hossain T, Anwar MN. Proteolytic activity of a *Lactobacillus species* isolated from rumen. Pakistan *Journal of Biological Science* 2004; 7(12): 2105-2108.
- 41. Raja MMM, Raja A, Sivasankari K et al. Production and characterization of protease enzyme isolated from *Penicillium sp* by solid state fermentation. *World Journal of Science and Technology* 2011; 1(9): 43-47.
- 42. Mishra A. Evaluation of the effect of Mutations on the Protease enzyme production by *Lactobacillus delbrueckii Helix* 2013; 5: 398-401.
- 43. Owoseni AA, Onilude AA. Characterization of Two Proteases from *Enterobacter* and *Escherichia coli* Isolated from Processed Foods in the West African Sub-Region. Report and Opinion 2012; 4(10): 48-53.