

Research Article

Expression of Apoptosis Activating and Cell Proliferation Proteins in Oral Lichen Planus

Navneet Saini¹, Rashmi Babbar², Ashish Kumar Mandal³

¹Assistant Professor, (PhD Physiology), Adesh Institute of Medical Sciences and Research, Punjab, India.

²Director and Professor, (MD Physiology), Maulana Azad Medical College, New Delhi, India.

³Professor, (MD Pathology), Maulana Azad Medical College, New Delhi, India.

DOI: <https://doi.org/10.24321/2394.6539.201905>

I N F O

Corresponding Author:

Navneet Saini, Adesh Institute of Medical Sciences and Research, Punjab, India.

E-mail Id:

drsaininavneet@yahoo.com

Orcid Id:

<https://orcid.org/0000-0002-1142-0604>

How to cite this article:

Saini N, Babbar R, Mandal AK. Expression of Apoptosis Activating and Cell Proliferation Proteins in Oral lichen Planus. J Adv Res Med Sci Tech. 2019;6(1&2)25-30.

Date of Submission: 2018-10-15

Date of Acceptance: 2019-01-10

A B S T R A C T

Aims and Objectives: Oral lichen planus is an immune-mediated mucocutaneous disease of unknown etiology characterized by the destruction of keratinocytes via apoptosis by infiltrating T lymphocytes. Though cytotoxic T lymphocytes can activate both extrinsic and intrinsic pathways of apoptosis, key proteins involved in both pathways have not been studied or there are conflicting results in english literature. The aim of the present paper was to study the key proteins involved in extrinsic & intrinsic pathways of apoptosis as well as products of genes involved in cell proliferation in cases of oral lichen planus.

Material and Methods: The study was undertaken at Maulana Azad Medical College New Delhi. Thirty histopathologically confirmed cases of oral lichen planus were included in this study. The presence of apoptosis was identified by TdT-mediated dUTP-biotin nick-end labelling. The expression of caspase-3, caspase-8, caspase-9, bax, K-ras, c-jun and c-erb-2 was studied by immunohistochemistry on histopathology sections.

Results: A statistically significant increase in all the proteins studied and apoptosis was found as compared to the control subjects. The association between apoptosis: caspase-3, caspase-3: capase-8, caspase-9: bax, caspase-8: caspase-9 and caspase-3: caspase-9 was also significant. An inverse association was found between c-jun: apoptosis, c-erb-2: apoptosis.

Conclusion: We found increased expression of proteins involved in both the pathways of apoptosis as well those associated with cell proliferation. Our results suggest that apoptosis is occurring via both pathways in oral lichen planus. There is also increased cell proliferation which may be responsible for the later development of malignancy in cases of oral lichen planus.

Keywords: Lichen, Apoptosis, Cytotoxic T-cells, Disease

Introduction

Lichen planus is a mucocutaneous disease, first described by Erasmus Wilson in 1869.¹ The prevalence of lichen planus ranges from 0.1%-2.2% worldwide.^{2,8} In the urban areas of India, the prevalence of oral lichen planus range from 0.02%-0.4%, while in villages it varies from 0.1%-1.5%.^{9,13}

The lichen planus is a disease of the middle-aged and elderly with most cases occurring from 30-70 years.^{14,16} However it has also been reported in infants under the age of six months.^{17,18} Bhattacharya et al. have reported the age range of 8-76 years in the Indian population.⁹

Its premalignant nature has been studied by many authors, who have reported a malignant transformation rate of up to 6.1%.^{19,35}

The etiology of lichen planus is unknown, and its association with tobacco intake is controversial.^{36,37} However oral lichen planus is now thought to be a cell-mediated immune response to an induced antigenic change in the mucosa, in which basal keratinocytes appear to be the target of T-lymphocytes.^{19,38}

The cytotoxic T-lymphocytes utilize various mechanisms involving the granzyme system,^{39,41} T-cell restricted intracellular antigen (Tia-1)⁴² and cytokines⁴³ to induce cell death. They also express Fas ligand on their surface and thus can stimulate death receptors present on the surface of keratinocytes.^{23,44,45} The cytotoxic-T-lymphocytes are also known to damage DNA (the exact mechanism of which is not clear), thereby activating the tumour suppressor p53 gene, which can either repair the damage by arresting the cell cycle or initiate the cell to undergo apoptosis.⁴⁶ Several pro-apoptotic members of Bcl-2 family like Bax, Bak and Apaf-1 (apoptosis activating factor) can be activated by p53 gene. These proteins further initiate the mitochondrial pathway of apoptosis.⁴⁷

An increase in apoptosis has been observed by many authors in oral lichen planus.^{48,50} The upregulation of bax and caspase-3 has also been reported by some authors,^{51,52,38} though Dekker et al⁴⁶ did not observe any significant increase in bax. We could not find any publication in the English literature on caspase-8 and caspase-9 in oral lichen planus.

In oral lichen planus enhanced expression of genes regulating cell proliferation has been reported by some workers.^{53,54} However their role has not been studied in detail, though some work on c-erb B-2 gene has been reported,^{53,54} K-ras and c-Jun expression have not been studied.

In the present study, we aimed to examine the expression of proteins activating two major pathways of apoptosis (death receptor and mitochondrial pathway) in oral lichen and their relation with proteins regulating cell proliferation.

Material and Methods

The study was undertaken at Maulana Azad Medical College New-Delhi. Thirty clinically diagnosed cases of oral lichen planus and thirty control subjects were included in the study. The oral biopsy specimen was taken from the representative area of each case of oral lichen planus. The diagnosis was confirmed by the presence of characteristic histopathological features of lichen planus. The control group included those subjects having non-specific mild clinical changes on oral mucosa and histology showing minimum inflammation in sub epithelium not specific for any pathology. The study was cleared by the ethical committee of the institute and informed consent was obtained from each subject. The expression of apoptosis, caspase-3, caspase-8, caspase-9, bax, K-ras, c-Jun and c-erbB-2 was studied in all the cases.

Apoptosis

The apoptotic cells were identified by in-situ Terminal TdT-mediated dUTP-biotin Nick-End Labelling (TUNEL) using a commercial kit (cat / q1A33, Oncogene, Boston, U.S.A). In brief, the tissue sections were deparaffinised, rehydrated and treated with Proteinase-K for 20 min at room temperature. The endogenous peroxidase was inactivated by 3% H₂O₂ in methanol. The tissue sections were incubated with TdT equilibration buffer for 30 min at room temperature, and then with labelling reaction mixture for 60 min at 37°C. The 50 X peroxide conjugate was diluted in the blocking buffer (1:50) and tissue sections were treated with it for 30 min at room temperature. All intermediate washing steps were performed by phosphate buffer saline (pH-7.4). The brown colour of the apoptotic cells was then developed by diaminobenzidine. Counterstaining was done with hematoxylin.

Immunohistochemistry

The caspase-8 (Novacastra, Newcastle, U.K), caspase-3 (Novacastra, Newcastle, U.K), caspase-9 (Novacastra, Newcastle, U.K), bax (Santa Cruze, California, U.S.A), K-ras (Santa Cruze, California, U.S.A), c-Jun (Novacastra, Newcastle, U.K) and c-erbB-2 (Novacastra, New castle, U.K) proteins were detected by immunostaining based on avidin-biotin peroxides complex technique.

The tissue section was deparaffinised and rehydrated. The endogenous peroxidase was inactivated with 3% H₂O₂ in methanol and non-specific protein binding was blocked by incubation with 5gm milk in 100 ml of phosphate buffer saline (pH-7.4). Slides were then incubated overnight with 40 µl of diluted primary antibody (20:1) at 4°C. Biotinylated horse antimouse IgG secondary antibodies were then applied for 30 min (Novacastra, Newcastle, U.K). The sections were incubated for 30 min with avidin-horse peroxides complex (Novacastra, Newcastle, U.K). All intermediate rinsing steps

were performed with phosphate buffer saline (pH-7). The brown colour was developed by diaminobenzidine. The positive expression of these antibodies was observed in the cytoplasm.

Statistical Analysis

Cases were divided into two categories: positive and negative depending on the expression of particular protein and apoptosis. The difference in the number of positive cases between the groups was studied by the chi-square test. If any of the expected frequencies was less than five then Fisher's exact test was applied. The association between the groups was studied by the chi-square test for association. A p-value of <0.05 was considered statistically significant.

Results and Observations

Immunohistochemical staining on 30 cases of oral lichen planus showed cytoplasmic positivity in 33 % (10 / 30) of cases for caspase-3 and in 53 % (16 / 30) for caspase-8. The caspase-9 positive cases were 53 % (16 / 30) and those for Bax protein were 67% (20 / 30). About 46 % (14 / 30) cases of lichen planus were positive for K-ras Table 1.

The membrane positivity for c-erb B-2 on co-protein was observed in 40 % (12 / 30) cases and c-Jun nuclear positivity was found in 33 % (10 / 30) cases. Keratinocytes in 40 % (12 / 30) cases of lichen planus specimens showed positive apoptotic signals. Table 1, Results of our control group are summarized in table 1, Figure 1.

A statistically significant increase in the positivity for these proteins and apoptosis was observed in lichen planus cases as compared to the control group Table 1.

We also observed a statistically significant association between apoptosis: caspase-3, caspase-3: caspase-8, caspase-9: bax, caspase-8: caspase-9 and caspase-9: caspase-3. The cases which were positive for apoptosis were also positive for caspases and/or bax, however inverse association was observed between c-Jun:apoptosis and c-erb- 2:apoptosis Table 2.

Table 1. The Percentage Positivity of Apoptosis Activating and Cell Proliferation Proteins

	Control Group (% Positivity) (n = 30)	Lichen Planus (% Positivity) (n = 30)	*P-value (Chi-Square Test)
Apoptosis	6.6 (n=2)	40 (n = 12)	0.005
Caspase-3	3.3 (n=1)	33 (n = 10)	0.005
Caspase-3	6.6 n=2)	53 n = 16)	<0.001
Caspase-3	0.0 (n=0)	53 (n = 16)	<0.001
Bax	3.3 (n = 1)	67 (n = 20)	<0.001

c-Jun	10 (n = 3)	33 (n = 10)	0.03
c-erb-2	10 (n = 3)	40 (n = 12)	0.009
K-ras	10 (n = 3)	46 (n = 14)	0.004

*All P-Values were significant

Table 2. Association between Markers of Apoptosis & Cell Proliferation in Oral Lichen Planus

Association Pair	P-value (Chi-Square Test)
Apoptosis	<0.001
Caspase-3	<0.001
Caspase-3	<0.001
Caspase-3	0.024
Bax	0.050
K-ras and apoptosis	0.300
c-Jun	0.024 [#]
c-erb-2	0.024 [#]
# inverse association	

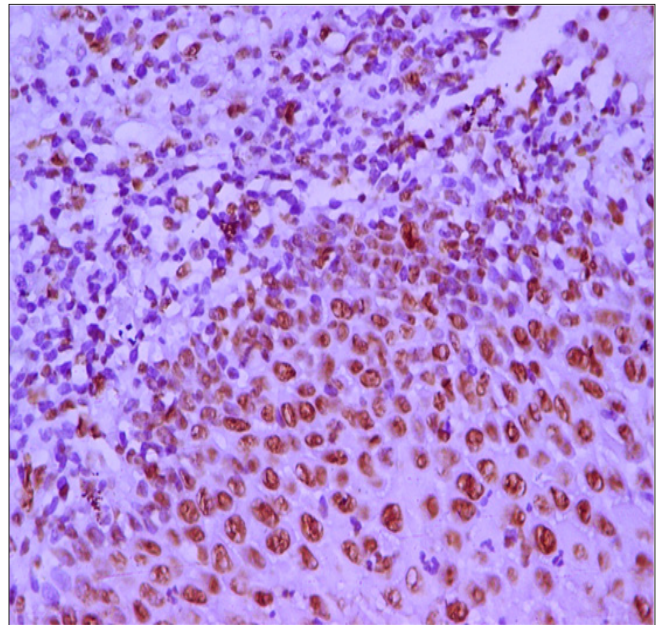


Figure 1. Expression of Apoptosis in Oral Lichen Planus

Discussion

Oral lichen planus is a chronic inflammatory disease, in which basal epithelial cells of the oral mucosa are attacked by T-lymphocytes.^{19,38} The target cells trigger a series of complex molecular mechanisms that may arrest the cell cycle for DNA repair, or induce apoptosis to eliminate cells with severely damaged DNA.⁴⁶ There are two major pathways by which the apoptotic demise of the cell is initiated, extrinsic pathway (death receptor) and intrinsic pathway (mitochondrial).^{23,47}

The activation of the Fas system on T-lymphocyte in oral lichen planus has been studied by various workers.^{23,44,45} The Fas ligand expressed by T-lymphocytes binds to the Fas receptor present on the surface of the epithelial cells. This results in the transduction of a signal that activates the death receptor pathway.^{23,44,45} We have studied the expression of proteins involved in this pathway in oral lichen planus. A significant increase in caspase-8 positivity was observed, similar increase was also observed in caspase-3 and apoptosis-positive cells as compared to the control subjects. Though we did not find any study of caspase-8 in oral lichen planus, the increase in caspase-3 and apoptosis positivity was also reported by other workers.^{48,51} The association of caspase-3 with caspase-8 and with apoptosis was significant in the present study, however contradictory results have been reported by some workers.⁴⁸ Thus our findings are consistent with the hypothesis that the apoptosis by death receptor pathway in oral lichen planus is being activated by fas ligand expressed on the cytotoxic T-lymphocytes.

In literature, the reports regarding bax positivity in oral lichen planus are conflicting.^{51,52,38,46} In the present study the up-regulation in Bax and caspase-9 has been observed. Some authors have suggested, that the increased Bax expression may be due to its activation by tumour suppressor p53 gene which gets activated due to DNA damage by cytotoxic T-lymphocytes.⁴⁶

The caspase-9 can also be activated by the death receptor pathway involving caspase-8, through pro-apoptotic molecule Bid.⁵⁵ A significant association of caspase-9 with caspase-8 in the present study may suggest the activation of this pathway by T-lymphocytes in oral lichen planus.

Thus we may summarize that cytotoxic T-lymphocytes can activate multiple pathways of apoptosis in oral lichen planus.

The substantial increase in apoptotic death of epithelial cells in oral lichen planus by cytotoxic T-lymphocyte could be harmful to the epithelium' however, some authors have demonstrated that epithelial cells in oral lichen planus frequently respond by evolving a resistance molecular mechanism that can resist the cell death and or stimulate the epithelial cell proliferation.^{53,54} Proto-oncogenes acting as growth factor receptors, signal transducers and transcription factors play a central role in controlling cell proliferation. A significant increase in c-erb B-2 (growth factor receptor), K-ras (signal transducer) and c-Jun(transcription factor) in the present study suggests the presence of a possible mechanism that might lead to the maintenance of the epithelial structure. The alteration in the expression of proto-oncogene regulating cell proliferation may lead to malignant transformation in oral lichen planus.

In conclusion, we found increased apoptosis in oral lichen planus. The apoptosis in oral lichen planus occurs via both extrinsic and intrinsic pathways, and the epithelium responds by increased expression of proteins involved in cell proliferation.

Source of Funding: None

Conflict of Interest: None

References

1. Wilson E. On lichen planus. J Cutan Med Dis Skin. 1869;3:117-32.
2. Meij EH, Schepman KP, Waal I. The possible premalignant character of oral lichen planus and oral lichenoid lesions: a prospective study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2003;96(2):164-71. [PubMed][Google Scholar]
3. Hellier FF. Environment and skin disease. Br J Dermatol. 1940;52:107-12. [Google Scholar]
4. Daoud MS, Pittelkow MR. Lichen planus. In: Goldsmith LA, Katz SI, Gilchrist BA, Paller AS, Leffell DJ, Wolff K, editors. Fitzpatrick's dermatology in general medicine. New York: McGraw, Hill Book Co. 2012;728.
5. White CJ. Lichen planus: a critical analysis of 64 cases. J Cutan Dis. 1919;37:639-42.
6. Samman PD. Lichen planus: an analysis of 200 cases. Trans St Johns Hosp Dermatol Soc. 1961;46:36-8. [PubMed][Google Scholar]
7. Altman J, Perry HO. The variants and course of lichen planus. Arch Dermatol. 1961;84:179-91. [PubMed][Google Scholar]
8. Sehgal VN, Rege VL. Lichen planus: an appraisal of 147 cases. Indian J Dermatol Venereol Leprol. 1974;48:104-7. [Google Scholar]
9. Bhattacharya M, Kaur I, Kumar B. Lichen planus: a clinical and epidemiological study. J Dermatol. 2000;27(9):576-82. [PubMed][Google Scholar]
10. Pindborg JJ, Chawla TN, Misra RK, Nagpaul RK, Gupta VK. Frequency of oral carcinoma, leukoplakia, leukokeratosis, leukoedema, submucous fibrosis and lichen planus in 10,000 Indians in Lucknow, Uttar Pradesh, India: preliminary report. J Dent Res. 1965;44:61. [PubMed][Google Scholar]
11. Zachariah J, Mathew B, Varma NA, Iqbal AM, Pindborg JJ. Frequency of oral mucosal lesions among 5000 individuals in Trivandrum, South India. Preliminary report. J Indian Dent Assoc. 1966;38(11):290-4. [PubMed]
12. Pindborg JJ, Mehta FS, Daftary DK, Gupta PC, Bhonsle RB. Prevalence of oral lichen planus among 7639 Indian villages in Kerala, South India. Acta Derm Venereol. 1972;52(3):216-20. [PubMed][Google Scholar]
13. Pindborg JJ, Kalapes HK, Kale SA, Singh B, Taleyerkhan BN. Frequency of oral leukoplakia and related conditions

- among 10,000 Bombayites. Preliminary report. *J All India Dent Assoc.* 1965;37(7):228-9. [PubMed]
14. Kovesi G, Banoczy J. Follow up studies in oral lichen planus. *IntJ Oral Surg.* 1973;2:13-7. [PubMed][Google Scholar]
 15. Lacy MF, Reade PC, Hay KD. Lichen planus: a theory of pathogenesis. *Oral Surg Oral Med Oral Pathol.* 1983;56:521-6. [PubMed][Google Scholar]
 16. Hallopeau H. Sur uncas de lichen de Wilson gingival avec neopla sievoisine dans la region maxillaire. *Bull Soc Fr Dermatol Syphiligr.* 1910;17:32. French.
 17. Culver GD. A clinical study of lichen planus. *Arch Dermatol Syph.* 1920;1:43-9. [Google Scholar]
 18. Pusey WA. Lichen planus in an infant less than six months old. *Arch Dermatol Syph.* 1929;19:671-2. [Google Scholar]
 19. Rajentheran R, McLean NR, Kelly CG, Reed MF, Nolan A. Malignant transformation of oral lichen planus. *Eur J Surg Oncol.* 1999;25:520-3. [PubMed][Google Scholar]
 20. Duffey DC, Eversole LR, Abemayor E. Oral lichen planus and its association with squamous cell carcinoma: an update on pathogenesis and treatment implications. *Laryngoscope.* 1996;106:357-62. [PubMed][Google Scholar]
 21. Markopoulos AK, Antoniadis D, Papanayotou P, Trigonidis G. Malignant potential of oral lichen planus: a follow-up study of 326 patients. *Oral Oncol.* 1997;33:263-9. [PubMed][Google Scholar]
 22. Fulling HJ. Cancer development in oral lichen planus: a follow-up study of 327 patients. *Arch Dermatol.* 1973;108:667-9. [PubMed][Google Scholar]
 23. Silverman Jr S, Gorsky M, Lozada-Nuf F. A prospective follow-up study of 570 patients with oral lichen planus: persistence, remission and malignant association. *Oral Surg Oral Med Oral Pathol.* 1985;60:30-4. [PubMed][Google Scholar]
 24. Murti PR, Daftary DK, Bhonsle RB, Gupta PC, Mehta FS, Pindborg JJ. Malignant potential of oral lichen planus observation in 722 patients from India. *J Oral Pathol.* 1986;15:71-7. [PubMed][Google Scholar]
 25. Holmstrup R, Thorn JJ, Rindum J, Pindborg JJ. Malignant development of lichen planus affected oral mucosa. *J Oral Pathol.* 1988;17:219-25. [PubMed][Google Scholar]
 26. Salem G. Oral lichen planus among 4277 patients from Gizan, Saudi Arabia. *Community Dent Oral Epidemiol.* 1989;17:322-4. [PubMed][Google Scholar]
 27. Silverman Jr S, Gorsky M, Lozada-Nur F, Giannotti K. A prospective study of findings and management in 214 patients with oral lichen planus. *Oral Surg Oral Med Oral Pathol.* 1991;72:665-70. [PubMed][Google Scholar]
 28. Sigurgeirsson B, Lindelof B. Lichen planus and malignancy: an epidemiologic study of 2071 patients and a review of the literature. *Arch Dermatol.* 1991;127:1684-8. [PubMed][Google Scholar]
 29. Voute AB, de Jong WF, Schulten EA, Snow GB, vander Wall I. Possible premalignant character of oral lichen planus: the Amsterdam experience. *J Oral Pathol Med.* 1992;21:326-9. [PubMed][Google Scholar]
 30. Brown RS, Bottomley WK, Puente E, Lavigne GL. A retrospective evaluation of 193 patients with oral lichen planus. *J Oral Pathol Med.* 1993;22:69-72. [PubMed][Google Scholar]
 31. Barnard NA, Scully C, Eveson JW, Cunningham S, Porter SR. Oral cancer development in patients with oral lichen planus. *J Oral Pathol Med.* 1993;22:421-4. [PubMed][Google Scholar]
 32. Silverman Jr S, Bahl S. Oral lichen planus update: clinical characteristics, treatment responses, and malignant transformation. *Am J Dent.* 1997;10(6):259-63. [PubMed][Google Scholar]
 33. Lo Muzio L, Mignogna MD, Favia G, Procaccini M, Testa NF, Bucci E. The possible association between oral lichen planus and oral squamous cell carcinoma: a clinical evaluation on 14 cases and a review of the literature. *Oral Oncol.* 1998;34:239-46. [PubMed][Google Scholar]
 34. Lanfranchi-Tizeira HE, Aguas SC, Sano SM. [Malignant transformation of atypical oral lichen planus: a review of 32 cases]. *Med Oral.* 2003;8:2-9. Spanish. [PubMed][Google Scholar]
 35. Bhonsle RB, Pindborg JJ, Gupta PC, Murti PR, Mehta FS. Incidence rate of oral lichen planus among Indian villages. *Acta Derm Venereol.* 1979;59:255-7. [PubMed][Google Scholar]
 36. Gupta PC, Murti PR, Bhonsle RB, Mehta FS, Pindborg JJ. Effect of cessation of tobacco use on the incidence of oral mucosal lesions in a 10-yr follow-up study of 12,212 users. *Oral Dis.* 1995;1:54-8. [PubMed][Google Scholar]
 37. Hedberg N, Ng A, Hunter N. A semi-quantitative assessment of the histopathology of oral lichen planus. *J Oral Pathol.* 1986;15:268-72. [PubMed][Google Scholar]
 38. Bloor BK, Malik FK, Odell EW, Morgan PR. Quantitative assessment of apoptosis in oral lichen planus. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 1999;88:187-95. [PubMed][Google Scholar]
 39. Tschopp J, Jongeneel ICV. Cytotoxic T lymphocyte mediated cytotoxicity. *Biochemistry.* 1988;27:2641-6. [PubMed][Google Scholar]
 40. Shi L, Kam CM, Powers JC, Aebbersold R, Greenberg AH. Purification of three cytotoxic lymphocyte granule

- serine proteases that induce apoptosis through distinct substrate and target cell interactions. *J Exp Med.* 1992;176:1521-9. [PubMed][Google Scholar]
41. Heusel JW, Wessel schmidt RL, Shresta S, Russell JH, Ley TJ. Cytotoxic lymphocytes require granzyme B for the rapid induction of DNA fragmentation and apoptosis in allogeneic target cells. *Cell.* 1994;76:977-87. [PubMed] [Google Scholar]
 42. Tian Q, Streuli M, Saito H, Schlossman SF, Anderson P. A polyadenylate binding protein localized to the granules of cytolytic lymphocytes induces DNA fragmentation in target cells. *Cell.* 1991;67:629-39. [PubMed][Google Scholar]
 43. Yamamoto T, Osaki T, Yoneda K, Ueta E. Cytokine production by keratinocytes and mononuclear infiltrates in oral lichen planus. *J Oral Pathol Med.* 1994;23:309-15. [PubMed][Google Scholar]
 44. Nagata S, Golstein P. The Fas death factor. *Science.* 1995;267:1449-53. [PubMed][Google Scholar]
 45. Quan LT, Tewari M, O'Rourke K, Dixit V, Snipas SJ, Poirier GG, Ray C, Pickup DJ, Salvesen GS. Proteolytic activation of the cell death protease Yama/CPP32 by granzyme B. *Proc Natl Acad Sci USA.* 1996;93:1972-6. [PubMed][Google Scholar]
 46. Dekker NP, Lozada-Nur F, Lagenaur LA, MacPhail LA, Bloom CY, Regezi JA. Apoptosis-associated markers in oral lichen planus. *J Oral Pathol Med.* 1997;26:170-5. [PubMed][Google Scholar]
 47. Reed JC. Double identity for proteins of the Bcl-2 family. *Nature.* 1997;387:773-6. [PubMed][Google Scholar]
 48. Tobon-Arroyave SI, Villegas-Acosta FA, Ruiz-Restrepo SM, Vieco-Duran B, Restrepo-Misas M, Londono-Lopez ML. Expression of caspase-3 and structural changes associated with apoptotic cell death of keratinocytes in oral lichen planus. *Oral Dis.* 2004;10:173-8. [PubMed] [Google Scholar]
 49. Karatsaidis A, Schreurs O, Axell T, Helgeland K, Schenck K. Identity of TUNEL- positive cells in the oral buccal epithelium of normal mucosa and lichen lesions. *J Oral Pathol Med.* 2004;33:264-8. [PubMed][Google Scholar]
 50. Tanda N, Mori S, Saito K, Ikawa K, Sakamoto S. Expression of apoptotic signaling proteins in leukoplakia and oral lichen planus: quantitative and topographical studies. *J Oral Pathol Med.* 2000;29:385-93. [PubMed] [Google Scholar]
 51. Bascones C, Gozalez-Moles MA, Esparza G, Bravo M, Acevedo A, Gil-Montoya JA, Bascones A. Apoptosis and cell cycle arrest in oral lichen planus: hypothesis on their possible influence on its malignant transformation. *Arch Oral Biol.* 2005;50:873-81. [PubMed][Google Scholar]
 52. Sklavovnou A, Chrysomali E, Scorilas A, Karameris A. TNF-alfa expression and apoptosis-regulating proteins in oral lichen planus: a comparative immunohistochemical evaluation. *J Oral Pathol Med.* 2000;29:370-5. [PubMed][Google Scholar]
 53. Kilpi A, Rich AM, Konttinen YT, Reade PC. The expression of c-erb B-2 protein in the keratinocytes of oral mucosal lichen planus. *Br J Dermatol.* 1995;133:847-52. [PubMed][Google Scholar]
 54. Taniguchi Y, Nagao T, Maeda H, Kameyama Y, Warnakulasuriya KA. Epithelial cell proliferation in oral lichen planus. *Cell Prolif.* 2002;35:103-9. [PubMed] [Google Scholar]
 55. Susin SA, Zamzami N, Castedo M, Hirsch T, Marchetti P, Macho A, Daugas E, Geukens M, Kroemer G. Bcl-2 inhibits the mitochondrial release of an apoptogenic protease. *J Exp Med.* 1996;184:1331-41. [PubMed] [Google Scholar]